

May 26, 1948.

Dear Ed-

Thanks for the information on Tergitol 7. Carbide & Carbon sent me an 8 oz. sample that should last the rest of my natural life. As soon as some long-awaited incubators come in, I'll get to work on indicator media, for Neurospora.

Here is a print from a cross of W-108 x Y-46 ( $\text{Lac}_3^- \times \text{f}^+$ ) on the best synthetic EMB we've hit so far. The formula is: (per liter):

Asparagine	10
Lactose	10
$(\text{NH}_4)_2\text{SO}_4$	5
NaCl	1
$\text{MgSO}_4$	.1
$\text{K}_2\text{HPO}_4$	2
Methylene Blue	.065
Eosin Y	.4
Agar	15

(In this case  $\text{B}_1$  also)

It works very well indeed to make up 10 or 20 liter batches in the form of a dry mix, and this is very much more convenient of course.

Aaron Novick, in Szilard's lab., has been working at getting recombination in K-12 mutants using phage, e.g. mixing 58-161/1/6 and W-1/4/7 and then selecting with T1, T6, T4, and T7. His early results indicate, if anything, a remarkably high yield of multiple resists in the mixed cultures, with satisfactory controls.

Anyhow, he finds that W-1 differs from K-12 in being sensitive to T3 (the other cultures are sensitive to a frequent mutant of T3) and I've promised to send him the intermediate cultures. However, I don't have 679, 679-680, and I wonder whether you could send me those at your convenience. Nothing else much; W-67 is behaving very peculiarly in its segregation for  $\text{Lac}_4^-$  and  $\text{V}_1^r$ , and may be an inversion. Esther's found what looks like suppressors of  $\text{Lac}_3^-$  as 'reversions' of W-112.

Best regards, and expect we'll be seeing you soon,